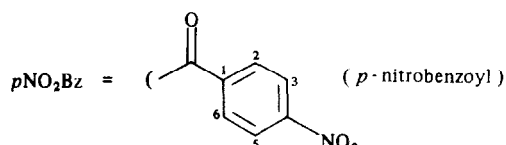
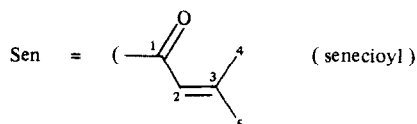
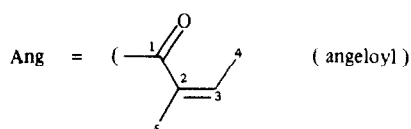


- 1** $R^1 = R^2 = H$
2 $R^1 = R^2 = \text{Ang}$
3 $R^1 = p\text{NO}_2\text{Bz}$, $R^2 = \text{OH}$
4 $R^1 = R^2 = p\text{NO}_2\text{Bz}$
5 $R^1 = p\text{NO}_2\text{Bz}$, $R^2 = \text{Sen}$
6 $R^1 = \text{OH}$, $R^2 = \text{Sen}$
7 $R^1 = \text{Ang}$, $R^2 = \text{Sen}$



(4.40), 248 (3.87); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1711 and 1653 ($\text{C}=\text{C}-\text{C}=\text{O}$, esters), 1672 and 1617 ($\text{C}=\text{C}-\text{C}=\text{O}$, ketone); ^1H NMR (300 MHz) and ^{13}C NMR (75.4 MHz): Tables 1 and 2 respectively.

Acknowledgements—We are grateful to Dr Jerzy Rzedowski for identifying the plant material, and to CoNaCyT and SEP-México for the financial support.

REFERENCES

- Gil, R. R., Oberti, J. C., Sosa, V. E. and Herz, W. (1987) *Phytochemistry* **26**, 1459.
- Amaro, J. M., Adrián, M., Cerda, C. M. and Joseph-Nathan, P. (1988) *Phytochemistry* **27**, 1409.
- Bohlmann, F., Ates, N., Jakupovic, J., King, R. M. and Robinson, H. (1982) *Phytochemistry* **21**, 2691.
- Bohlmann, F., Suwita, A., Natu, A. A., Czerson, H. and Suwita, A. (1977) *Chem. Ber.* **110**, 3572.
- Román, L. U., del Río, R. E., Hernández, J. D., Cerda, C. M., Cervantes, D., Castañeda, R. and Joseph-Nathan, P. (1985) *J. Org. Chem.* **50**, 3965.
- Bohlmann, F. and Zdero, C. (1985) *Liebigs Ann. Chem.* 1764.
- Nayak, U. R. and Dev, S. (1968) *Tetrahedron* **24**, 4099.
- Beeby, P. J. (1977) *Tetrahedron Letters* 3379.
- Kindt-Larsen, T., Bitsch, V., Andersen, I. G. K., Jart, A. and Munch-Petersen, J. (1963) *Acta Chem. Scand.* **17**, 1426.
- Bohlmann, F., and Tietze, B. M. (1970) *Chem. Ber.* **103**, 561.
- Joseph-Nathan, P., Cerda, C. M., del Río, R. E., Román, L. U. and Hernández, J. D. (1986) *J. Nat. Prod.* **49**, 1053.

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SESQUITERPENE LACTONES FROM *PYRETHRUM SANTOLINOIDES*

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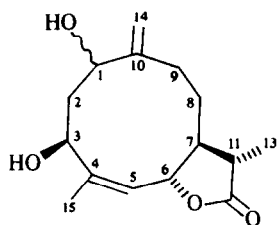
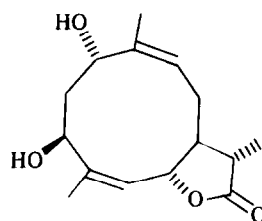
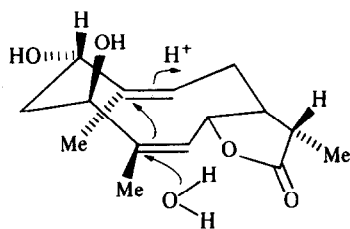
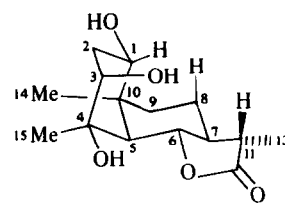
Key Word Index—*Pyrethrum santolinoides*; Compositae; sesquiterpene lactones; germacranolides; heliangolides; eudesmanolide.

Abstract—A reinvestigation of the aerial parts of *Pyrethrum santolinoides* afforded, in addition to known compounds, four new sesquiterpene lactones, three germacranolides and an eudesmanolide. The structures were elucidated by high field ^1H NMR techniques.

INTRODUCTION

Pyrethrum santolinoides DC (= *Tanacetum sinaicum* Del. ex DC) has been studied previously. From the aerial parts

some sesquiterpene lactones were reported [1] and from the roots various compounds, including unusual triterpenes, were isolated [2]. We have studied a sample of the aerial parts collected at Wadi Elarbaeen, Sanct Kathe-

**1** 1 α OH**2** 1 β OH**3****4** 9 β ,10 β epoxide**3****5**

rine, Sinai peninsula and the results are discussed in this paper.

RESULTS AND DISCUSSION

The extract of the aerial parts afforded, in addition to some widespread compounds (see Experimental), the

germacranolide **1**, which has been isolated previously from the species [1], and three further ones, the epimeric diol **2**, the Δ^9 -isomer **3** and the corresponding epoxide **4**. Furthermore the eudesmanolide **5** was present.

The structure of compound **2** followed from its ^1H NMR spectrum (Table 1) which was in part similar to that of **1**. However, several signals were shifted and also

Table 1. ^1H NMR spectral data of compounds **2**–**5** (400 MHz, CDCl_3 , δ -values)

H	2	3	4	5
1	4.10 <i>br ddd</i>	5.03 <i>dd</i>	3.92 <i>dd</i>	3.73 <i>br dd</i>
2 α	2.34 <i>ddd</i>	1.92 <i>ddd</i>	2.16 <i>m</i>	1.82 <i>ddd</i>
2 β	2.28 <i>ddd</i>	2.16 <i>ddd</i>	2.25 <i>m</i>	2.44 <i>ddd</i>
3	4.46 <i>br dd</i>	4.48 <i>dd</i>	4.48 <i>br s</i>	3.15 <i>br dd</i>
5	5.24 <i>dq</i>	5.09 <i>br d</i>	5.18 <i>br d</i>	1.83 <i>d</i>
6	5.88 <i>dd</i>	5.94 <i>dd</i>	5.94 <i>dd</i>	3.98 <i>dd</i>
7	1.81 <i>m</i>	1.47 <i>dddd</i>	1.75 <i>br ddd</i>	1.60 <i>m</i>
8 α	1.76 <i>m</i>	2.22 <i>m</i>	2.36 <i>br d</i>	1.75 <i>m</i>
8 β	2.08 <i>m</i>	2.45 <i>ddd</i>	1.50 <i>ddd</i>	1.27 <i>m</i>
9 α	2.23 <i>br dd</i>			1.08 <i>ddd</i>
9 β	2.62 <i>br dd</i>	5.23 <i>ddq</i>	2.83 <i>dd</i>	2.26 <i>m</i>
11	2.35 <i>m</i>	2.24 <i>m</i>	2.31 <i>dq</i>	2.26 <i>m</i>
13	1.24 <i>d</i>	1.22 <i>d</i>	1.26 <i>d</i>	1.23 <i>d</i>
14	5.32 <i>br s</i>	1.68 <i>dd</i>	1.24 <i>s</i>	0.97 <i>s</i>
15	5.13 <i>br s</i>			
	1.78 <i>d</i>	1.67 <i>d</i>	1.80 <i>br s</i>	1.43 <i>s</i>

$J[\text{Hz}]$: compound **2**: 1,2 α = 4; 1,2 β = 10; 2 α ,2 β = 16; 2 α ,3 = 5; 2 β ,3 = 4; 3, OH = 6; 5,6 = 6,7 = 10; 5,15 = 1.5; 8 β ,9 = 10; 8 α ,9 α = 8; 9 α ,9 β = 15; 11,13 = 7; compound **3**: 1,2 β = 11.5; 1,2 α = 2; 2 α ,2 β = 14.5; 2 β ,3 = 2.5; 2 α ,3 = 5; 5,6 = 6,7 = 10; 5,15 = 1.5; 7,8 β = 8 β ,9 = 11; 7,8 α = 2; 7,11 = 12; 8 α ,8 β = 13.5; 8 α ,9 = 6; 8 β ,14 = 9,14 = 1; 11,13 = 7; compound **4**: 1,2 β = 10.5; 1,2 α = 2.5; 5,6 = 6,7 = 10; 7,8 β = 8 β ,9 = 11; 7,11 = 12; 8 α ,8 β = 14.5; 8 α ,9 = 3; 11,13 = 7; compound **5**: 1,2 β = 6; 1,2 α = 10; 2 α ,2 β = 15; 2 β ,3 = 1.5; 2 α ,3 = 2; 5,6 = 6,7 = 10; 8 α ,9 β = 4; 8 β ,9 β = 9 α ,9 β = 13.5; 11,13 = 7.

some of the couplings differed. Spin decoupling indicated that only the configuration at C-1 was changed in the lactone. In agreement with the down field shift of H-14 the lactone **2** had a 1β -hydroxy group.

The molecular formula of compound **3** ($C_{15}H_{22}O_4$) showed that we were dealing with an isomer of compounds **1** and **2**. The 1H NMR spectrum (Table 1) indicated that the exomethylene group was replaced by an olefinic methyl group. Accordingly, two signals (δ 1.68 *dd* and 1.67 *d*) for olefinic methyls were visible and in addition to a broadened doublet at δ 5.09 (5-H) a further low field signal (δ 5.23 *ddq*) was present. Spin decoupling allowed the assignment of all signals, only those of H-8 α and H-11 being overlapped. The configuration at C-11 could not be deduced directly from the 1H NMR spectrum and the preferred conformation had to be determined for a clear assignment of the configurations. This was achieved by NOE difference spectroscopy. The observed effects required the proposed configurations and a conformation with both methyls below the plane but not perpendicular (Table 2). The NOE's between H-15 and H-5 as well as between H-9 and H-14 established the presence of a heliangolide with a Δ^9 -Z-configuration for the double bond. This germacranolide type is relatively rare, for example some exomethylene lactones with 8β -acyloxy groups of this type have been reported from *Eupatorium* species [3].

In the 1H NMR spectrum of **4** (Table 1) one of the olefinic proton signals was replaced by a double doublet at δ 2.83. As followed from spin decoupling this signal was due to H-9. The chemical shift required the presence of a proton at an epoxide carbon. Thus, this lactone was the epoxide of **3**. This was established by determining the stereochemistry by the NOE which requires a conformation as in the case of **3** with a β -epoxide moiety. Inspection of models clearly showed that an α -epoxide can not have a conformation which would agree with the observed NOE's and couplings. Compounds **1-4** are probably all derived from a 3β -hydroxyheliangolide with a 1(10)-double bond by attack of oxygen accompanied by allylic rearrangement and epoxidation in the case of **4**.

The 1H NMR spectral data of **5** (Table 1) indicated that an eudesmanolide was present. The signals were in part similar to those of reynosin. However, the exomethylene proton signals were replaced by a sharp singlet at δ 1.43.

Accordingly, a tertiary methyl carbinol was very likely. A double doublet at δ 3.15 indicated a further hydroxy group and a methyl doublet at δ 1.23 agreed with the presence of a 11,13-dihydro lactone. Spin decoupling showed that the signal of the corresponding vicinal proton (H-11) was overlapped by a multiplet. Therefore, no clear coupling with H-7 could be recognized. However, NOE difference spectroscopy allowed the assignment of the stereochemistry at C-11 and also at all other chiral centres. A clear effect between H-1, H-6 and H-8 β as well as a NOE of H-14 with H-5 required the presence of a *cis*-decalin system (Table 2). Most likely the lactone **5** is formed by cyclization of **3** as shown in Scheme 1. Inspection of a model of **3** shows that in the preferred conformation it would lead to a *cis*-decalin system with a stereochemistry as observed for lactone **5**.

EXPERIMENTAL

The air-dried plant material (640 g) was extracted with Et_2O -MeOH-petrol (1:1:1) for 24 hr at room temp. After defatting with MeOH the extract was separated first by CC (silica gel) as reported previously [4] into three fractions (Fr. 1: Et_2O -petrol, 1:9 and 1:3; Fr. 2: Et_2O -petrol, 1:1 and Fr. 3: Et_2O and Et_2O -MeOH, 9:1). TLC of fraction 1 gave 50 mg thymol, of fraction 2 gave 5 mg thymol, 15 mg *p*-methoxyacetophenone and 23 mg stigmasterol. TLC of one-sixth of fraction 3 (Et_2O -MeOH, 99:1) gave two bands (Fr. 3/1 and Fr. 3/2). TLC and HPLC of Fr. 3/1 (MeOH- H_2O , 7:3, always RP 18, flow rate 3 ml/min) gave 3 mg **5** (R_f , 4.8 min). HPLC of Fr. 3/2 (MeOH- H_2O , 3:1) gave 3 mg **4** (R_f , 1.0 min), 2.5 mg **2** (R_f , 1.4 min), 3 mg **1** (R_f , 2.1 min) and 7 mg **3** (R_f , 2.9 min). Known compounds were identified by comparing the 400 MHz 1H NMR spectra with those of authentic material.

1 α ,3 β -Dihydroxy-7 α ,11 β H-germacra-4Z,10(14)-dien-12,6 α -olide (**2**). Colourless gum; IR $\nu_{max}^{CHCl_3}$ cm^{-1} : 3600 (OH), 1770 (γ -lactone); CIMS m/z (rel. int.): 267 [$M+1$] $^+$ (92) ($C_{15}H_{22}O_4+1$), 249 [267- H_2O] $^+$ (100), 231 [249- H_2O] $^+$ (48); [α] $_D^{24}$ +46° ($CHCl_3$; *c* 0.18).

1 α ,3 β -Dihydroxy-7 α ,11 β H-germacra-4Z,9Z-dien-12,6 α -olide (**3**). Colourless crystals, mp. 163°; IR $\nu_{max}^{CHCl_3}$ cm^{-1} : 3600 (OH), 1770 (γ -lactone); MS m/z (rel. int.): 266.152 [M] $^+$ (14) (calc. for $C_{15}H_{22}O_4$: 266.152), 248 [$M-H_2O$] $^+$ (14), 233 [248-Me] $^+$ (9), 222 [$M-CO_2$] $^+$ (12), 100 (100); [α] $_D^{24}$ +154° ($CHCl_3$; *c* 0.13).

Table 2. NOE's of the lactones **3-5** (in parenthesis % NOE)

Irradiated	3	4	5
H-1	H-6 H-8 β (3) (4)	H-6 H-8 β (3) (3)	H-6 H-8 β (10) (5)
H-3	H-2 α H-2 β H-15 (5) (3) (5)	H-2 α H-2 β H-15 (6) (6) (5)	H-2 α H-2 β H-15 (3) (6) (14)
H-5	H-15 H-7 (5) (3)	H-7 H-15 (3) (4)	H-14 H-15
H-6	H-1 H-8 β H-11 (4) (2) (3)	H-1 H-8 β H-11 (3) (2) (5)	H-1 H-11 H-8 β (8) (7) (3)
H-9 (9°)	H-14 H-7 (5) (3)	H-7 H-14 (3) (2)	H-14 (4)
H-14	H-9 α (10)	H-9 α H-15 (10) (6)	H-2 α H-5 H-9 α H-15 (5) (10) (5) (2)
H-15	H-3 H-5 (7) (9)	H-3 H-5 H-14 (5) (8) (4)	H-3 H-5 H-14 (7) (5) (2)

1 α ,3 β -Dihydroxy-9 β ,10 β -epoxy-7 α ,11 β H-germacra-4Z-en-12,6 α -olide (4). Colourless gum; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3600 (OH), 1770 (γ -lactone); MS m/z (rel. int.): 266.152 $[\text{M} - \text{H}_2\text{O}]^+$ (1.5) (calc. for $\text{C}_{15}\text{H}_{22}\text{O}_4$; 266.152), 251 $[\text{266} - \text{Me}]^+$ (6), 109 (70), 95 (97), 55 (100); $[\alpha]_{\text{D}}^{24} + 34^\circ$ (CHCl_3 ; c 0.21).

1 α ,3 β ,4 β -Trihydroxy-(5 α ,7 α ,11 β H-10 α methyl)-eudesman-12,6 α -olide (5). Colourless gum; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3600 (OH), 1770 (γ -lactone); MS m/z (rel. int.): 266.152 $[\text{M} - \text{H}_2\text{O}]^+$ (6) (calc. for $\text{C}_{15}\text{H}_{22}\text{O}_4$; 266.152), 251 $[\text{266} - \text{Me}]^+$ (11), 248 $[\text{266} - \text{H}_2\text{O}]^+$ (7), 233 $[\text{251} - \text{H}_2\text{O}]^+$ (5), 192 (100); $[\alpha]_{\text{D}}^{24} + 10^\circ$ (CHCl_3 ; c 0.33).

REFERENCES

1. El-Sebakhy, N. A., El-Ghazouly, M. G., Seif El-Din, A. A. and Zdero, C. (1986) *Pharmazie* **41**, 525.
2. Jakupovic, J., Eid, F., Bohlmann, F. and El-Dahmy, S. (1987) *Phytochemistry* **26**, 1536.
3. Herz, W., de Groote, R. and Murari, R. (1978) *J. Org. Chem.* **43**, 3559.
4. Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1984) *Phytochemistry* **23**, 1979.

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ONOSERIOLIDE DERIVATIVES FROM *LEUCERIA* SPECIES

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Key Word Index—*Leuceria leontopodioides*, *L. purpurea*, *L. thermanum*, *L. hahnii*; Compositae; sesquiterpene lactones; onoseriolides; acetylenic compounds.

Abstract—From *Leuceria* species in addition to known, widespread compounds, four new onoseriolide derivatives and a known tetraynene together with an isomeric derivative, were isolated. The structures were elucidated by high field NMR techniques. The chemotaxonomy is discussed briefly.

INTRODUCTION

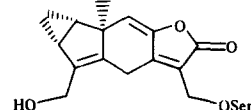
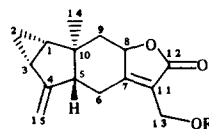
The genus *Leuceria* has 46 species distributed from south Peru to Patagonia [1] but so far only one species has been studied chemically. In addition to the tetrayne derivative **6** the rare sesquiterpene lactone onoseriolide (**4**) and its Δ^8 -derivative were isolated [2]. We have studied four further species and the results are discussed in this paper.

RESULTS AND DISCUSSION

The aerial parts of *L. leontopodioides* (O. K.) Schum. afforded in addition to curcumene and tridecapentaynene, the onoseriolide derivatives **1**–**3** and **5**, and the tetraynes **6** [3] and **7**. From the extract of *L. purpurea* (Vahl.) H. et A. also tridecapentaynene and the lactone **3** were isolated while the aerial parts of *L. thermanum* (Phil.) Phil. only gave triterpenes (see Experimental) and those of *L. hahnii* Franchet gave lupenone, tridecapentaynene and tetradeca-4,6-dien-8,10,12-triyn-1-ol.

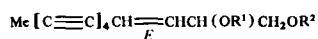
The structures of **1** and **2** were deduced from the ^1H NMR spectra (Table 1) as they were similar to those of onoseriolide (**4**) and its 8,9-dihydro derivative [4, 5] respectively. However, the typical signals of senecioates and the down field shift of the H-13 signals indicated that the corresponding ester were present. The ^1H NMR

spectrum of **3** (Table 1) was similar to that of **2**. However, the low field singlet at δ 6.43 was replaced by a singlet at δ 4.21. As followed from the molecular formula ($\text{C}_{20}\text{H}_{22}\text{O}_5$) lactone **3** had one more oxygen. Accordingly, all data agreed with the presence of the epoxide of compound **2**. This was supported by the ^{13}C NMR data (Table 2) especially if compared with those of **1** and **2** (Table 2). The configuration at C-8 and C-9 could not be determined. However, biogenetic considerations strongly favour a β -epoxidation of the Δ^8 -bond. The ^1H NMR spectrum of **5** (Table 1) was in part similar to that of compound **2**. As already followed from the molecular



- 1** R = Sen, **2** R = Sen, Δ^8
3 R = Sen, 8 β , 9 β -epoxide
4 R = H

5



- 6** $\text{R}^1 = \text{H}$, $\text{R}^2 = i\text{Bu}$ **7** $\text{R}^1 = i\text{Bu}$, $\text{R}^2 = \text{H}$